Supplementary Material



Supplemental Figure S1. Gating strategy of vAT flow cytometry analysis. (**A**) The gating strategy of the flow cytometry analysis of vAT. First, immune cells are selected based on CD45 and side scatter (SSC). From this subset, B-cells, NK-cells, T-cells and granulocytes are removed using B220, NK, CD3 and Ly6G. Cells negative for this cocktail are selected and used to determine F4/80 and CD11c expression. The cells represented in gate 1 are F4/80⁺CD11c⁻ M2 macrophages and can be subdivided into F4/80⁺CD11c⁻MHC^{high} and F4/80⁺CD11c⁻MHC^{low}. The cells represented in gate 2 are F4/80^{low}CD11c⁺ and can be divided into F4/80, CD11c (**B**), CD11b (**C**) and MHC2 (**D**) staining.



Supplemental Figure S2. F4/80^{high}CD11c⁻ MHC2^{low} macrophages are bona fide phagocytosing macrophages. LDLR^{-/-} mice were fed a HFD and two days prior to sacrifice, clodronate liposomes are injected to deplete macrophages (n = 3/group). Uptake of the clodronate liposomes results in depletion of F4/80^{high}CD11c⁻MHC2^{low} (A), F4/80^{high}CD11c⁻MHC2^{high} (B) and F4/80^{int}CD11c⁺CD11b^{high} (C) ATMs whereas the dendritic cells (F4/80⁻CD11c⁺CD11b^{low} (D) remain unaffected showing that F4/80^{high}CD11c⁻ MHC2^{low} cells are indeed phagocytosing macrophages. Data is presented as mean ± SEM and analyzed using unpaired students t-test. * p<0.05



Supplemental Figure S3. HFD induced changes in the immune cell populations in the vAT. LDLR^{-/-} mice were fed a HFD or control diet for either 5 weeks or 16 weeks (n = 7/group). Flow cytometry was used to determine the various immune cell subsets in the vAT: CD45⁺ immune cells, monocytes, dendritic cells (DC), NK cells, B-cell, CD4⁺ T-cells, CD8⁺ T-cells and granulocytes. Data is presented as mean ± SEM and analyzed using one-way ANOVA and Tukey's multiple comparison test. ** p<0.01, ***p<0.001



Supplemental Figure S4. HFD induced changes in the immune cell populations in the subcutaneous (sc)AT. LDLR^{-/-} mice were fed a HFD or control diet for either 5 weeks or 16 weeks (n = 7/group). Flow cytometry was used to determine the various immune cell subsets in the scAT: CD45⁺ immune cells, monocytes, dendritic cells (DC), F4/80lowCD11c⁺ M1 macrophages, F4/80⁺CD11c⁻MHC2^{low} M2 macrophage subset, F4/80⁺CD11c⁻MHC2^{high} M2 macrophage subset, granulocytes, B-cell, CD8⁺ T-cells, CD4⁺ T-cells and NK cells. Data is presented as mean ± SEM and analyzed using one-way ANOVA and Tukey's multiple comparison test. *p<0.05, ** p<0.01, ***p<0.001, ****p<0.001

Supplemental Table S1. The total amount of cells sorted from the vAT of chow (sample 1 – 3) and HFD (sample 4 – 6) fed LDLR^{-/-} **mice for RNA extraction.** Sample 1 – 3 are a pool of n=6 LDLR^{-/-} mice/sample and sample 4 – 6 are a pool of n=4 LDLR^{-/-} mice/sample. After sorting, these samples are used for RNA extraction and microarray analysis.

	CD11c⁺	CD11c ⁻ MHC2 ^{low}	CD11c ⁻ MHC2 ^{high}
Sample 1	80.138	94.041	218.870
Sample 2	52.989	68.183	138.003
Sample 3	97.592	129.999	403.988
Sample 4	609.583	218.746	274.308
Sample 5	671.260	271.292	302.414
Sample 6	567.515	151.871	268.821

Supplemental Table S2. Microarray data from the cluster mobility of CD11c⁺, CD11c⁻MHC2^{low} and CD11c⁻MHC2^{high} ATM sorted from vAT of LDLR^{-/-} mice after HFD. Microarray data showed the fold change and adjusted p-value of the various genes represented in the cluster mobility.

		CD11c ⁺		CD11c ⁻ MHC2 ^{low}		CD11c ⁻ MHC2 ^{high}	
Gene	Description	Fold change	Adjusted p-value	Fold change	Adjuste d p- value	Fold change	Adjuste d p- value
Ccl4	chemokine (C-C motif) ligand 4	1.033614	0.952041	2.65275	0.00318	1.75455	0.09067
						4	5
Ccr2	chemokine (C-C motif) receptor	-1.44551	0.062303	2.33281	0.00030	1.39017	0.16136
	2			7	6		9
Ccr5	chemokine (C-C motif) receptor	1.090284	0.806116	1.70569	0.04407	1.30706	0.43145
	5			8	3	5	1
Cxcr4	chemokine (C-X-C motif)	1.536505	0.107202	1.88158	0.03541	1.43743	0.29684
	receptor 4			8	2	1	4
Pdgfb	platelet derived growth factor, B	2.133762	0.020665	2.43347	0.01655	1.74966	0.15470
	polypeptide			4	7	4	4
Myo1f	myosin IF	1.44276	0.039203	1.65258	0.01178	1.37646	0.12071
					5	5	
Tgfbr1	transforming growth factor,	2.651582	0.001036	1.93594	0.04395	1.71295	0.09458
	beta receptor I			9	1	8	2

		CD11c ⁺		CD11c ⁻ MHC2 ^{low}		CD11c ⁻ MHC2 ^{high}	
Gene	Description	Fold change	Adjuste d p- value	Fold change	Adjusted p-value	Fold change	Adjuste d p- value
H2-Aa	histocompatibility 2, class II	1 02826	0.90779	1.79337	0.005541	-	0.99658
	antigen A, alpha	-1.03030	9	8		1.00491	5
H2-Ab1	histocompatibility 2, class II	-1.10103	0.71386	1.77360	0.006394	1.00554	0.99564
	antigen A, beta 1		4	6		8	9
H2-	histocompatibility 2, class II,	1 2216	0.43795	1 0/865	0.009922	-	0.96838
DMa	locus DMa	-1.2310	1	1.54005		1.03947	6
H2-	histocompatibility 2, class II,	1 1969	0.76476	2.00945	0.021756	1.05277	0.96245
DMb1	locus Mb1	-1.1202	3	5		8	6
H2-Eb1	histocompatibility 2, class II	-1.07124	0.86859	2.49481	0.001416	-1.0093	0.99481
	antigen E beta		4	3			1
Ciita	class II transactivator	-1.23128	0.51893	2.02500	0.018145	-	0.97189
			4	3		1.04206	1
Fcgr4	Fc receptor, IgG, low affinity IV	1.998848	0.14399	3.17541	0.027762	2.46842	0.08299
			6	9		4	9

Supplemental Table S3. Microarray data from the cluster immune response of CD11c⁺, CD11c⁻MHC2^{low} and CD11c⁻MHC2^{high} ATM sorted from vAT of LDLR^{-/-} mice after HFD. Microarray data showed the fold change and adjusted p-value of the various genes represented in the cluster immune response.